

宫颈癌放射治疗中DNA损伤修复相关基因

罗 然, 李贤富*, 陈俊安, 刘文君

川北医学院附属医院放疗科, 四川省肿瘤学重点学科, 医学影像四川省重点实验室, 四川 南充
Email: 175708021722@qq.com, *lixianfu13@163.com

收稿日期: 2020年12月14日; 录用日期: 2021年1月3日; 发布日期: 2021年1月18日

摘 要

宫颈癌是全世界女性最常见的恶性肿瘤之一, 也是女性癌症死亡的第二大常见原因。放射治疗可在宫颈癌患者的所有阶段进行。辐射抗性是肿瘤放射治疗中的关键问题。因此, 研究宫颈癌放射治疗过程中DNA损伤修复相关基因, 有利于寻找新的治疗靶点以进一步提高宫颈癌放疗疗效。

关键词

宫颈癌, 放射治疗, DNA损伤修复, *GS*, *LINP1*, *VRK1*, *HOTAIR*

DNA Damage Repair Related Genes in Cervical Cancer Radiotherapy

Ran Luo, Xianfu Li*, Jun'an Chen, Wenjun Liu

Key Laboratory of Medical Imaging of Sichuan Province, Key Discipline of Oncology of Sichuan Province, Department of Radiotherapy, Affiliated Hospital of North Sichuan Medical College, Nanchong Sichuan
Email: 175708021722@qq.com, *lixianfu13@163.com

Received: Dec. 14th, 2020; accepted: Jan. 3rd, 2021; published: Jan. 18th, 2021

Abstract

Cervical cancer is one of the most common malignant tumors in women around the world, and it is also the second most common cause of cancer death in women. Radiation therapy can be performed at all stages of cervical cancer patients. Radiation resistance is a key issue in tumor radiotherapy. Therefore, the study of genes related to DNA damage and repair in the course of cervical cancer radiotherapy is helpful to find new therapeutic targets to improve further cervical cancer radiotherapy efficacy.

*通讯作者。

文章引用: 罗然, 李贤富, 陈俊安, 刘文君. 宫颈癌放射治疗中DNA损伤修复相关基因[J]. 临床医学进展, 2021, 11(1): 132-137. DOI: 10.12677/acm.2021.111019

Keywords

Cervical Cancer, Radiation Therapy, DNA Damage and Repair, *GS*, *LINP1*, *VRK1*, *HOTAIR*

Copyright © 2021 by author(s) and Hans Publishers Inc.

This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

1. 前言

宫颈癌是全世界女性最常见的恶性肿瘤之一[1]。虽然现在已经有了人类乳头状瘤病毒(HPV)疫苗,并且已经建立了有效的筛查方法来降低其发病率和死亡率,但宫颈癌仍然是发展中国家女性中第二常见的癌症和第三主要的癌症死亡原因[2]。2016年根据国家癌症综合网络(National Comprehensive Cancer Network, NCCN)指南指出,放射治疗可在宫颈癌患者的所有阶段进行,包括:作为无法手术的早期患者的治疗;在较大的早期和晚期肿瘤中联合化疗或作为术后的辅助治疗;作为姑息治疗,可缓解癌症引起的症状并改善晚期患者的生活质量。但是,有些患者对放射治疗的反应较差,复发和治疗失败的发生率较高[3] [4] [5]。

放射抗性是肿瘤放射治疗中的关键问题。放疗引起DNA损伤进而杀死肿瘤细胞,特别是DNA双链断裂(DSBs)。DNA双链断裂本身通常不是致命的,但它使细胞无法修复DNA双链断裂最终导致细胞死亡[6]。因此,DNA损伤修复能力是影响细胞放射反应的重要因素之一[7]。因此,关于哪些因素是宫颈癌的抗放射性和不良预后的原因,还有待于更多研究。

真核生物中两种主要的DNA损伤修复途径是同源重组(HR)和非同源末端连接(NHEJ) [8]。这些修复系统在亚致死性损伤和产生剂量率效应中起重要作用,如果损伤得以完整和精确的修复,细胞的放射敏感性就会发生改变;如果损伤不能被修复,则会诱导细胞凋亡[9]。本文将对宫颈癌放射治疗DNA损伤修复相关基因进行综述。

2. 谷氨酰胺合成酶(*GS*)

谷氨酰胺合成酶(*GS*)是一种代谢酶,它催化谷氨酸和氨的连接形成谷氨酰胺[10]。肿瘤细胞利用谷氨酰胺作为必需的代谢原料,以满足肿瘤细胞快速增殖的能量需求[11] [12] [13]。耐辐射细胞具有高谷氨酰胺合成代谢。因此,谷氨酰胺及其代谢底物可为DNA损伤修复提供调控机制[14]。*GS*的下调可延迟DNA修复,减弱核苷酸代谢,增强体内外的辐射敏感性;*GS*的高表达促使肿瘤细胞在放射时仍能生长。Fu S.等人[6]研究在辐射诱导应激下,谷氨酰胺代谢对肿瘤放射治疗反应的影响。他们发现抗辐射细胞具有较高的谷氨酰胺合成代谢。该研究在体外实验中发现*GS*基因敲除可延迟DNA修复,减弱核苷酸代谢,增强放射敏感性。最终结果显示谷氨酰胺合成酶通过促进核苷酸合成和加速DNA修复,将谷氨酰胺代谢与放疗反应联系起来。也有研究证实谷氨酰胺合成酶与胰腺癌、前列腺癌、胃癌、肺癌等肿瘤辐射抗性有关[15] [16] [17] [18],需要更进一步研究谷氨酰胺合成酶与宫颈癌辐射抗性的关系。

3. 非同源末端连接途径1 (*LINP1*)

非同源末端连接被认为是哺乳动物细胞DSB修复的主要途径。此种途径的修复过程是在不需要DNA末端之间广泛的同源性连接实现的[19]。研究显示*LINP1*促进非同源末端连接介导的DNA损伤修复活性[20] [21],其表达可能受EGFR途径调控[22]。Liang等[23]人研究表明非同源末端连接途径1 (*LINP1*)与乳腺癌细胞的增殖、转移和化学抗性有关。Li等[24]人通过qRT-PCR和Western blot方法检测,探讨*LINP1*

在卵巢癌中的作用机制, 结果发现卵巢癌组织中 *LINPI* 的表达明显高于癌旁组织, 并证实 *LINPI* 通过下调 *KLF6* 促进肿瘤细胞的转移和增殖。Wang X.等[25]人使用 γ -H2AX 聚焦形成实验观察放疗前后宫颈癌 HeLa S3 细胞的凋亡情况, 发现 *LINPI* 通过 NHEJ 途径提高 DNA 损伤修复效率, 参与宫颈癌的抗辐射性。虽然 *LINPI* 的表达水平与宫颈癌患者放疗后肿瘤反应的关系尚未完全阐明, 但他们的研究表明 *LINPI* 可能是宫颈癌的一个重要的预后指标和一个新的治疗靶点。

4. *VRK1*

越来越多的证据表明, 长链非编码 RNA (*lncRNAs*) 在妇科肿瘤的凋亡、转移、侵袭、迁移和细胞增殖中起着重要的生理作用[26], *VRK1* 是其中一种。*VRK1* [25] [27] 是一种丝氨酸激酶, 与细胞增殖[28] 和 DNA 损伤反应有关[29] [30] [31]。Li 等人[32] 用 qPCR 检测乳腺癌细胞中 *VRK1* 的表达情况, 发现 *VRK1* 在乳腺癌组织中表达下调, 抑制细胞增殖, 促进细胞凋亡, 与肿瘤分期降低、生存期延长有关。*VRK1* 在细胞增殖和 DNA 修复过程中起着至关重要的作用, 如果被抑制, 可以导致肿瘤生长减缓, 基因不稳定性增加, 也可能使肿瘤更具免疫原性, 可用于治疗[33]。*VRK1* 在宫颈癌细胞 DNA 放射损伤修复中的作用机制需进一步研究探讨。

5. *HOTAIR*(长非编码核糖核酸 HOX 转录反义 RNA)

HOX 转录反义 RNA (*HOTAIR*) 是一个 2158 核苷酸长的 *lncRNA*, 从染色体 12q13.13 上的 HOXC 位点表达, 它被发现能够沉默肿瘤抑制因子, 激活癌基因和关键信号通路[34] [35] [36] [37] [38]。体内实验还发现, *HOTAIR* 在宫颈癌组织中高表达, 并与临床分期、肿瘤大小和淋巴结转移相关[39] [40]。Guo X. 等[41] 人研究发现 *HOTAIR* 的功能被抑制可以通过抑制宫颈癌 HeLa 细胞的 Wnt 信号通路, 减少自噬和逆转上皮-间充质转化, 从而提高对放疗的敏感性。也有研究表明 *HOTAIR* 的敲除可通过抑制 *Wnt*/ β -*catenin* 途径抑制细胞增殖并在 G1 期阻滞[42]。此外, Jing L. 等人[43] 的研究证明 *HOTAIR* 的敲除上调了 *p21* 的表达, 从而提高了宫颈癌细胞的放射敏感性。持续稳定的 *HOTAIR* 基因敲除显著抑制肿瘤生长, 并使宫颈癌对体内放疗敏感。他们的研究提供了新的证据, 说明 *HOTAIR* 可以促进宫颈癌的增殖、侵袭和转移。这些发现表明, 靶向 *HOTAIR* 治疗宫颈癌是一种有效的治疗策略。

6. 研究展望

恶性肿瘤治疗的一个主要方法是在肿瘤细胞中产生灾难性的 DNA 损伤, 目前放射治疗是恶性肿瘤治疗的常用方法, 其能直接损伤肿瘤细胞 DNA。放射治疗可在宫颈癌治疗的所有阶段进行, 如根治性放疗、术前或术后辅助放疗及晚期或复发病例。尽管宫颈癌放疗已取得显著效果, 现在也使用铂类等放疗增敏剂, 但仍存在一些放疗效果不佳或复发病例。在放射治疗过程中, DNA 双链断裂未及时得到修复, 即可引起肿瘤细胞凋亡。肿瘤细胞的抗辐射性一直是限制宫颈癌治疗效果和导致其复发的主要障碍。目前关于宫颈癌放射性损伤 DNA 修复相关基因的研究是热点, 如 *GS*、*LINPI*、*VRK1*、*HOTAIR* 等。因此需要更多试验去研究宫颈癌放疗过程中 DNA 损伤修复相关基因及其机制, 从而发现一些新的治疗策略以提高宫颈癌放疗疗效。

基金项目

四川省教育厅自然科学重点项目(17ZA0173), 四川卫生计生委项目(19PJ037), 南充市市校合作科研专项(18SXHZ0398)。

参考文献

- [1] Tan, L.T., Tanderup, K., Kirisits, C., de Leeuw, A., Nout, R., Duke, S., et al. (2019) Image-Guided Adaptive Radiotherapy in Cervical Cancer. *Seminars in Radiation Oncology*, 29, 284-298.

- <https://doi.org/10.1016/j.semradonc.2019.02.010>
- [2] Torre, L.A., Bray, F., Siegel, R.L., Ferlay, J., Lortet-Tieulent, J. and Jemal, A. (2015) Global Cancer Statistics, 2012. *CA: A Cancer Journal for Clinicians*, **65**, 87-108. <https://doi.org/10.3322/caac.21262>
 - [3] Hong, J.H., Tsai, C.S., Wang, C.C., Lai, C.H., Chen, W.C., Lee, S.P., *et al.* (2000) Comparison of Clinical Behaviors and Responses to Radiation between Squamous Cell Carcinomas and Adenocarcinomas/Adenosquamous Carcinomas of the Cervix. *Chang Gung Medical Journal*, **23**, 396-404.
 - [4] Zhou, J., Wu, S.G., Sun, J.Y., Li, F.Y., Lin, H.X., Chen, Q.H., *et al.* (2017) Comparison of Clinical Outcomes of Squamous Cell Carcinoma, Adenocarcinoma, and Adenosquamous Carcinoma of the Uterine Cervix after Definitive Radiotherapy: A Population-Based Analysis. *Journal of Cancer Research and Clinical Oncology*, **143**, 115-122. <https://doi.org/10.1007/s00432-016-2246-9>
 - [5] Chen, J.L., Huang, C.Y., Huang, Y.S., Chen, R.J., Wang, C.W., Chen, Y.H., *et al.* (2014) Differential Clinical Characteristics, Treatment Response and Prognosis of Locally Advanced Adenocarcinoma/Adenosquamous Carcinoma and Squamous Cell Carcinoma of Cervix Treated with Definitive Radiotherapy. *Acta Obstetrica et Gynecologica Scandinavica*, **93**, 661-668. <https://doi.org/10.1111/aogs.12383>
 - [6] Fu, S., Li, Z., Xiao, L.B., Hu, W.F., Zhan, L., Xie, B.W., *et al.* (2019) Glutamine Synthetase Promotes Radiation Resistance via Facilitating Nucleotide Metabolism and Subsequent DNA Damage Repair. *Cell Reports*, **28**, 1136-1143.e4. <https://doi.org/10.1016/j.celrep.2019.07.002>
 - [7] Nambiar, M. and Raghavan, S.C. (2011) How Dose DNA Break during Chromosomal Translocations? *Nucleic Acids Research*, **39**, 5813-5825. <https://doi.org/10.1093/nar/gkr223>
 - [8] Pardo, B., Go'mez-Gonza'lez, B. and Aguilera, A. (2009) DNA Repair in Mammalian Cells: DNA Double-Strand Break Repair: How to Fix a Broken Relationship. *Cellular and Molecular Life Sciences*, **66**, 1039-1056. <https://doi.org/10.1007/s00018-009-8740-3>
 - [9] 王济东, 王俊杰. 低剂量率辐射生物效应的研究进展[J]. 国外医学(放射医学核医学分册), 2005, 29(4): 186-189. <http://dx.chinadoi.cn/10.3760/cma.j.issn.1673-4114.2005.04.013>
 - [10] Krajewski, W.W., Collins, R., Holmberg-Schiavone, L., Jones, T.A., Karlberg, T. and Mowbray, S.L. (2008) Crystal Structures of Mammalian Glutamine Synthetases Illustrate Substrate-Induced Conformational Changes and Provide Opportunities for Drug and Herbicide Design. *Journal of Molecular Biology*, **375**, 217-228. <https://doi.org/10.1016/j.jmb.2007.10.029>
 - [11] Tardito, S., Oudin, A., Ahmed, S.U., Fack, F., Keunen, O., Zheng, L., Miletic, H., *et al.* (2015) Glutamine Synthetase Activity Fuels Nucleotide Biosynthesis and Supports Growth of Glutamine-Restricted Glioblastoma. *Nature Cell Biology*, **17**, 1556-1568. <https://doi.org/10.1038/ncb3272>
 - [12] Bott Alex, J., Peng, I.C., Fan, Y., Faubert, B., Zhao, L., Li, J., Neidler, S., Sun, Y., Jaber, N., Krokowski, D., *et al.* (2015) Oncogenic Myc Induces Expression of Glutamine Synthetase through Promoter Demethylation. *Cell Metabolism*, **22**, 1068-1077. <https://doi.org/10.1016/j.cmet.2015.09.025>
 - [13] Fluteau, A., Ince, P.G., Minett, T., Matthews, F.E., Brayne, C., Garwood, C.J., Ratcliffe, L.E., Morgan, S., Heath, P.R., Shaw, P.J., *et al.* (2015) The Nuclear Retention of Transcription Factor FOXO3a Correlates with a DNA Damage Response and Increased Glutamine Synthetase Expression by Astrocytes Suggesting a Neuroprotective Role in the Ageing Brain. *Neuroscience Letters*, **609**, 11-17. <https://doi.org/10.1016/j.neulet.2015.10.001>
 - [14] Turgeon, M., Perry, N.J.S. and Poulogiannis, G. (2018) DNA Damage, Repair, and Cancer Metabolism. *Frontiers in Oncology*, **8**, 15. <https://doi.org/10.3389/fonc.2018.00015>
 - [15] Wang, L., Peng, W., Wu, T.M., Deng, P.C. and Zhao, Y.L. (2018) Increased Glutamine Anabolism Sensitizes Non-Small Cell Lung Cancer to Gefitinib Treatment. *Cell Death Discovery*, **4**, Article No. 84. <https://doi.org/10.1038/s41420-018-0086-x>
 - [16] Ye, J.X., Huang, Q., Xu, J., Huang, J.S., Wang, J.Z., Zhong, W.J., *et al.* (2018) Targeting of Glutamine Transporter ASCT2 and Glutamine Synthetase Suppresses Gastric Cancer Cell Growth. *Journal of Cancer Research and Clinical Oncology*, **144**, 821-833. <https://doi.org/10.1007/s00432-018-2605-9>
 - [17] Li, D.D., Fu, Z.Q., Chen, R.W., Zha, X.H., Zhou, Y., Zeng, B., *et al.* (2015) Inhibition of Glutamine Metabolism Counteracts Pancreatic Cancer Stem Cell Features and Sensitizes Cells to Radiotherapy. *Oncotarget*, **6**, 31151-31163. <https://doi.org/10.18632/oncotarget.5150>
 - [18] Shi, X.C., Zhang, X.S., Yi, C., Liu, Y. and Qiao, H. (2014) [¹³N]Ammonia Positron Emission Tomographic/Computed Tomographic Imaging Targeting Glutamine Synthetase Expression in Prostate Cancer. *Molecular Imaging*, **13**, 1-10. <https://doi.org/10.2310/7290.2014.00048>
 - [19] Karran, P. (2000) DNA Double Strand Break Repair in Mammalian Cells. *Current Opinion in Genetics & Development*, **10**, 144-150. [https://doi.org/10.1016/S0959-437X\(00\)00069-1](https://doi.org/10.1016/S0959-437X(00)00069-1)
 - [20] Zhang, Y.Y., He, Q., Hu, Z.Y., Feng, Y., Fan, L.L., Tang, Z.Q., *et al.* (2016) Long Noncoding RNA *LINPI* Regulates

- Repair of DNA Double-Strand Breaks in Triple-Negative Breast Cancer. *Nature Structural & Molecular Biology*, **23**, 522-530. <https://doi.org/10.1038/nsmb.3211>
- [21] Zlotorynski, E. (2016) Non-Coding RNA: *LINP1* Joins Ends with Triple-Negative Effect. *Nature Reviews Molecular Cell Biology*, **17**, 330-331. <https://doi.org/10.1038/nrm.2016.60>
- [22] Wang, X.X., Liu, H., Shi, L.M., Yu, X.L., Gu, Y.J. and Sun, X.N. (2018) *LINP1* Facilitates DNA Damage Repair through Non-Homologous End Joining (NHEJ) Pathway and Subsequently Decreases the Sensitivity of Cervical Cancer Cells to Ionizing Radiation. *Cell Cycle*, **17**, 439-447. <https://doi.org/10.1080/15384101.2018.1442625>
- [23] Liang, Y.R., Li, Y.M., Song, X.J., Zhang, N., Sang, Y.T., Zhang, H.W., *et al.* (2018) Long Noncoding RNA *LINP1* Acts as an Oncogene and Promotes Chemoresistance in Breast Cancer. *Cancer Biology & Therapy*, **19**, 120-131. <https://doi.org/10.1080/15384047.2017.1394543>
- [24] Li, Y., Hou, C.Z., Dong, Y.-L., Zhu, L. and Xu, H. (2020) Long Noncoding RNA *LINP1* Promoted Proliferation and Invasion of Ovarian Cancer via Inhibiting KLF6. *European Review for Medical and Pharmacological Sciences*, **24**, 7918. https://doi.org/10.26355/eurrev_202008_22452
- [25] Campillo-Marcos, I. and Lazo, P.A. (2018) Implication of the *VRK1* Chromatin Kinase in the Signaling Responses to DNA Damage: A Therapeutic Target? *Cellular and Molecular Life Sciences*, **75**, 2375-2388. <https://doi.org/10.1007/s00018-018-2811-2>
- [26] Hosseini, E.S., Meryet-Figuire, M., Sabzalipoor, H., Haddad Kashani, H., Nikzad, H. and Asemi, Z. (2017) Dysregulated Expression of Long Noncoding RNAs in Gynecologic Cancers. *Molecular Cancer*, **16**, Article No. 107. <https://doi.org/10.1186/s12943-017-0671-2>
- [27] Cantarero, L., Moura, D.S., Salzano, M., Monsalve, D.M., Campillo-Marcos, I., Martín-Doncel, E. and Lazo, P.A. (2017) *VRK1* (Vaccinia-Related Kinase 1). In: Choi, S., Ed., *Encyclopedia of Signaling Molecules*, 2nd Edition, Springer Science, New York. https://doi.org/10.1007/978-1-4614-6438-9_561-2
- [28] Valbuena, A., Sanz-Garcia, M., Lopez-Sanchez, I., Vega, F.M., Lazo, P.A. (2011) Roles of *VRK1* as a New Player in the Control of Biological Processes Required for Cell Division. *Cellular Signalling*, **23**, 1267-1272. <https://doi.org/10.1016/j.cellsig.2011.04.002>
- [29] Sanz-Garcia, M., Monsalve, D.M., Sevilla, A. and Lazo, P.A. (2012) Vaccinia-Related Kinase 1 (*VRK1*) is an Upstream Nucleosomal Kinase Required for the Assembly of 53BP1 Foci in Response to Ionizing Radiation-Induced DNA Damage. *The Journal of Biological Chemistry*, **287**, 23757-23768. <https://doi.org/10.1074/jbc.M112.353102>
- [30] Monsalve, D.M., Campillo-Marcos, I., Salzano, M., Sanz-Garcia, M., Cantarero, L., Lazo, P.A. (2016) *VRK1* Phosphorylates and Protects NBS1 from Ubiquitination and Proteasomal Degradation in Response to DNA Damage. *Biochimica et Biophysica Acta (BBA)—Molecular Cell Research*, **1863**, 760-769. <https://doi.org/10.1016/j.bbamcr.2016.02.005>
- [31] Salzano, M., Sanz-Garcia, M., Monsalve, D.M., Moura, D.S. and Lazo, P.A. (2015) *VRK1* Chromatin Kinase Phosphorylates H2AX and Is Required for Foci Formation Induced by DNA Damage. *Epigenetics*, **10**, 373-383. <https://doi.org/10.1080/15592294.2015.1028708>
- [32] Li, Yang and Li, H. (2020) Circular RNA *VRK1* Correlates with Favourable Prognosis, Inhibits Cell Proliferation but Promotes Apoptosis in Breast Cancer. *Journal of Clinical Laboratory Analysis*, **34**, e22980. <https://doi.org/10.1002/jcla.22980>
- [33] Campillo-Marcos, I. and Lazo, P.A. (2019) Olaparib and Ionizing Radiation Trigger a Cooperative DNA-Damage Repair Response That Is Impaired by Depletion of the *VRK1* Chromatin Kinase. *Journal of Experimental & Clinical Cancer Research*, **38**, Article No. 203. <https://doi.org/10.1186/s13046-019-1204-1>
- [34] Carrion, K., Dyo, J., Patel, V., Sasik, R., Mohamed, S.A., Hardiman, G., *et al.* (2014) The Long Non-Coding *HOTAIR* is Modulated by Cyclic Stretch and WNT/beta-CATENIN in Human Aortic Valve Cells and Is a Novel Repressor of Calcification Genes. *PLoS ONE*, **9**, e96577. <https://doi.org/10.1371/journal.pone.0096577>
- [35] Yang, G., Zhang, S.H., Gao, F., Liu, Z.Y., Lu, M.J., Peng, S., *et al.* (2014) Osteopontin Enhances the Expression of *HOTAIR* in Cancer Cells via IRF1. *Biochimica et Biophysica Acta (BBA)—Gene Regulatory Mechanisms*, **1839**, 837-848. <https://doi.org/10.1016/j.bbagr.2014.06.020>
- [36] Zhang, H.Y., Cai, K., Wang, J., Wang, X.Y., Cheng, K., Shi, F.F., *et al.* (2014) MiR-7, Inhibited Indirectly by LincRNA *HOTAIR*, Directly Inhibits SETDB1 and Reverses the EMT of Breast Cancer Stem Cells by Downregulating the STAT3 Pathway. *Stem Cells*, **32**, 2858-2868.
- [37] Qiu, J.J., Lin, Y.Y., Ye, L.C., Ding, J.X., Feng, W.W., Jin, H.Y., *et al.* (2014) Overexpression of Long Non-Coding RNA *HOTAIR* Predicts Poor Patient Prognosis and Promotes Tumormetastasis in Epithelial Ovarian Cancer. *Gynecologic Oncology*, **134**, 121-128. <https://doi.org/10.1016/j.ygyno.2014.03.556>
- [38] Liu, X.H., Sun, M., Nie, F.Q., Ge, Y.B., Zhang, E.B., Yin, D.D., *et al.* (2014) Lnc RNA *HOTAIR* Functions as a Com-

-
- peting Endogenous RNA to Regulate HER2 Expression by Sponging miR-331-3p in Gastric Cancer. *Molecular Cancer*, **13**, Article No. 92. <https://doi.org/10.1186/1476-4598-13-92>
- [39] Prensner, J.R. and Chinnaiyan, A.M. (2011) The Emergence of lncRNAs in Cancer Biology. *Cancer Discovery*, **1**, 391-407. <https://doi.org/10.1158/2159-8290.CD-11-0209>
- [40] Prensner, J.R., Iyer, M.K., Alejandro Balbin, O., Dhanasekaran, S.M., Cao, Q., Brenner, J.C., *et al.* (2011) Transcriptome Sequencing across a Prostate Cancer Cohort Identifies PCAT-1, an Unannotated lncRNA Implicated in Disease Progression. *Nature Biotechnology*, **29**, 742-749. <https://doi.org/10.1038/nbt.1914>
- [41] Guo, X.G., Xiao, H.Q., Guo, S.H., Li, J., Wang, Y.X., Chen, J., *et al.* (2019) Long Noncoding RNA *HOTAIR* Knockdown Inhibits Autophagy and Epithelial-Mesenchymal Transition through the Wnt Signaling Pathway in Radioresistant Human Cervical Cancer HeLa Cells. *Journal of Cellular Physiology*, **234**, 3478-3489. <https://doi.org/10.1002/jcp.26828>
- [42] Li, J., Yang, S.Q., Su, N., Wang, Y., Yu, J.J., Qiu, H.F., *et al.* (2016) Overexpression of Long Non-Coding RNA *HOTAIR* Leads to Chemoresistance by Activating the Wnt/ β -Catenin Pathway in Human Ovarian Cancer. *Tumour Biology*, **37**, 2057-2065. <https://doi.org/10.1007/s13277-015-3998-6>
- [43] Li, J., Wang, Y., Dong, R.F., Yu, J.J. and Qiu, H.F. (2015) *HOTAIR* Enhanced Aggressive Biological Behaviors and Induced Radio-Resistance via Inhibiting p21 in Cervical Cancer. *Tumor Biology*, **36**, 3611-3619. <https://doi.org/10.1007/s13277-014-2998-2>