

款冬花化学成分的研究

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摘要

目的: 研究款冬花中的化学成分。方法: 综合应用反相色谱、Sephadex LH-20、液相和半制备液相等多种化学色谱方法进行分离纯化, 并根据化合物的理化性质和核磁共振波谱进行结构鉴定。结果: 从款冬花的95%乙醇提取物中分离得到20个化合物, 通过波谱分析等手段鉴定了15个化合物, 包括12个倍半萜, 2个脂肪酸和1个生物碱。结论: 化合物6和15为首次从款冬花中分离得到的。

关键词

款冬花, 倍半萜, 化学成分

Study on Chemical Constituents of *Tussilago farfara* L.

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Abstract

Objective: To study the constituents of *Tussilago farfara* L. **Methods:** The compounds were isolated and purified by column chromatography of Sephadex LH-20, HPLC and semi-preparative RP-HPLC. Their structures were elucidated by physicochemical properties and spectral analyses. **Results:** By the means of chromatography methods and spectroscopic evidence, 20 compounds were isolated from 95% EtOH of *T. farfara*. We identified 15 compounds by spectroscopic analysis, 12 of which were sesquiterpenoids. **Conclusion:** Compounds 6 and 15 are identified from *Tussilago farfara* L. for the first time.

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Keywords

Tussilago farfara L., Sesquiterpenoids, Chemical Constituents

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1. 引言

倍半萜类化合物由三个异戊二烯单位构成，含有 15 个碳原子。主要分布在植物界和微生物界，多以挥发油形式存在。这些年，对此类化合物的研究较快，每年发现的新型种类数目成倍增长，无论是化合物的数目还是骨架类型都是萜类化合物中最多的一类[1]。倍半萜类广泛存在于菊科植物中，并表现出有趣的化学多样性和重要的生物学特性，这使它们成为植物化学、药理学，及合成的主要目标[2]。

款冬花系菊科款冬属植物款冬(*Tussilago farfara* L.)的花蕾，是一种多年生草本植物，通常在 10 月下旬至 12 月下旬花尚未出土时采挖，主要分布于中国，欧洲和北非。款冬花中含有最多的成分是倍半萜，也是化学和药理研究最深入的一部分。根据其中所含有的倍半萜母核，主要以 oplopane 骨架和 bisabolane 骨架为主。款冬花中已报道的 oplopane 型倍半萜共有四十多个，占已报道倍半萜的一半以上。此类倍半萜的不同之处在于其取代基的位置变化，结构变化主要发生在 1、7 和 14 位。款冬花中报道的倍半萜中也有少部分是 bisabolane 型，该类倍半萜的结构特点是在 1、5 位和 8 位容易氧化，1 位和 8 位大多酰化成酯。近年来，也有学者从款冬花植物中发现了两个新颖结构的 oplopane 型倍半萜和一个新颖的 bisabolane 型倍半萜[3]。除了 oplopane 型和 bisabolane 型倍半萜外，款冬花中还报道了 eudesmane 型[4]倍半萜和 guaiane 型[5]倍半萜。我们对在陕西药材市场采购的款冬花进行了化学成分研究，从中分离得到了 20 个化合物，通过波谱分析手段最终鉴定了 15 个化合物(如图 1)，分别为：6,10-Octadecadienoic acid (1) [6]、Hexadecanoic acid (2) [7]、Senkirkine (3) [8]、(-)-cryptomerion (4) [9]、2,2-dimethyl-6-acetylchromanone (5) [10]、 β -oplophenone (6) [11]、7 β -(3-ethyl-*cis*-crotonoyloxy)-14-hydroxy-notonipetranone (7) [12]、14-acetoxy-7 β -(3-ethyl-*cis*-crotonoyloxy)-1 α -(2-methylbutyryloxy)-notonipetranone (8) [12]、tussilagone (9) [13]、7 β -(3-ethyl-*cis*-crotonoyloxy)-1 α -(2-methylbutyryloxy)-3(14)-dehydro-Z-notonipetranone (10) [13]、(1R,3R,4R,5S,6S)-1,5-diacetoxy-8-angeloyloxy-3,4-epoxybisabola-7(14),10-dien-2-one (11) [14]、(1R,3R,4R,5S,6S)-1-acetoxy-8-angeloyloxy-3,4-epoxy-5-hydroxybisabola-7(14),10-dien-2-one (12) [14]、1 β ,8-bisangeloyloxy-3 α ,4 α -epoxybisabola-7(14),10-dien-2-one (13) [3]、1 β -(3-ethyl-*cis*-crotonoyloxy)-8-angeloyloxy-3 α ,4 α -epoxybisabola-7(14),10-dien-2-one (14) [3]、(4R,6E)-2-acetoxy-8-angeloyloxy-4-hydroxybisabola-2,6,10-trien-1-one (15) [15]。其中，化合物 6 和 15 为首次从款冬花中分离得到的。

2. 仪器与试剂

1D 和 2D NMR 在 Bruker AM-400、DRX-500 或 Bruker AM-600 核磁共振仪上测定，TMS 作为内标， δ 为 ppm, J 为 Hz; ESI-MS 在 Burker HCT 或 Esquire 质谱仪上测定；高分辨质谱在 Auto-Spec Premier P776 质谱仪上测定；拌样及层析用硅胶(100~200, 200~300 目)，均为青岛海洋化工厂生产；反相填充材料 RP-18 为 40~60 μm ，Merk 公司生产；MCI 填充材料为 MCI-gel CHP-20P；HPLC 分析仪器为 Agilent 1260 型高效液相色谱仪，色谱柱为 Agilent 公司的 ZORBAX SB-C18 反相柱。凝胶为 Sephadex LH-20；显色剂非碱

为 10% H₂SO₄ 的乙醇溶液，喷洒后适当加热；生物碱常用显色剂：Dragendorff's，沾湿后显色。

3. 提取与分离

干燥款冬花 10 kg，粉碎后用 95% 的乙醇提取三次(25 L/次)，合并提取液，减压蒸馏除去有机溶剂，用乙酸乙酯进行萃取，得到粗提物 143.7 g，以聚酰胺拌样，以甲醇水系统(30%~100%)在 MCI 柱上进行梯度洗脱，以 TLC 进行检测，合并相同组分得到三个馏分：I-III。对萃取后的水层进行酸碱处理，酸层用乙酸乙酯萃取三次，得水层进行碱处理，并用氯仿萃取三次，得到生物碱 5.3 g。

馏分 I 段有部分结晶，得到化合物 2 (1.5 g)。

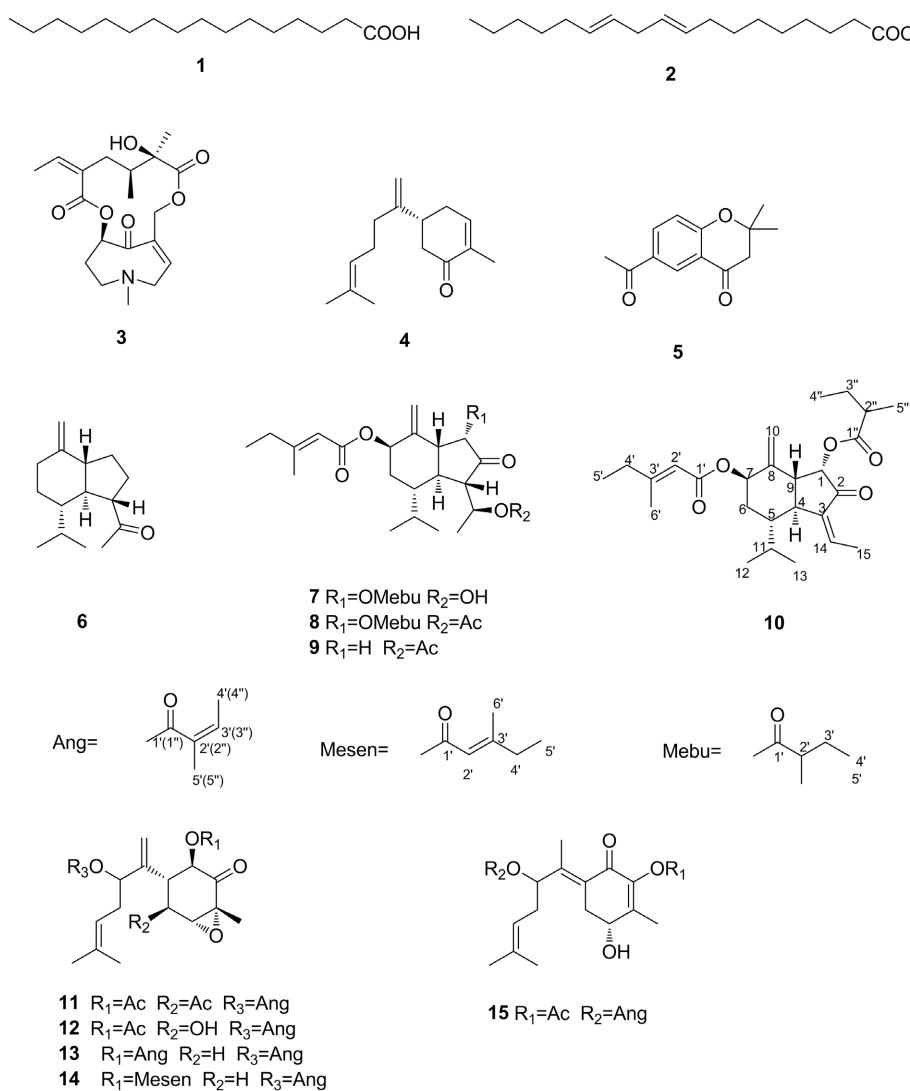


Figure 1. Obtained structural expressions of 15 compounds
图 1. 分离得到 15 个化合物的结构式

馏分 II 段有小部分结晶，得到化合物 9 (2.4 g)。将未结晶的部分，以聚酰胺拌样，以甲醇 - 水系统 (75%~100%) 在 RP-18 柱上进行梯度洗脱，以 TLC 进行检测，合并相同组分得到 I-a、I-b、I-c、I-d 和 I-e。I-a 经过多次硅胶柱层析后得到化合物 10 (2.3 g)，4 (4.1 mg) 和 1 (28.9 mg)。以甲醇-水系统(82: 28，流速

3 mL/min)在分析型 HPLC 对 I-b 段进行半制备后得到化合物 14 (20.0 mg, tR = 12.6 min), 13 (20.0 mg, tR = 11.2 min), 和 8 (3.8 mg, tR = 15.1 min)。I-c 以氯仿为洗脱剂进行柱层析后得到化合物 5 (65.2 mg)和混合物, 以甲醇-水系统(70:30, 流速 4 mL/min)在分析型 HPLC 对混合物进行半制备后得到化合物 11 (10.6 mg, tR = 11.9 min)。I-c 经过多次硅胶柱层析和半制备后得到化合物 6 (27.3 mg)和 7 (43.2 mg, tR = 11.8 min)。对 I-e 段进行半制备(甲醇-水 75:25, 流速 4 mL/min)得到化合物 12 (22.2 mg, tR = 10.8 min)和 15 (3.4 mg, 15.3 min)

生物碱部分经过多次硅胶柱层析后得到化合物 3 (1.1 g)。

4. 结构鉴定

6,10-Octadecadienoic acid (1): 无色油状物, $C_{18}H_{32}O_2$ 1H NMR (400 MHz, $CDCl_3$) δ_H : 6~5.35 (4H, m, H-9, 10, 12, 13), 2.77 (2H, t, J = 6.1 Hz, H-11), 2.34 (2H, t, J = 7.4 Hz, H-2), 2.05 (4H, m, H-8, 14), 1.63 (2H, m, H-3), 1.31 (14H, brs, H-4~7, H-15~17), 0.89 (3H, t, J = 6.7 Hz, H-18); ^{13}C NMR (100 MHz, $CDCl_3$) δ_C : 14.0 (C-18, CH_3), 22.6 (CH_2), 24.6 (CH_2), 25.6 (CH_2), 27.1 (CH_2), 27.2 (CH_2), 29.0 (CH_2), 29.0 (CH_2), 29.1 (CH_2), 29.3(CH_2), 29.6 (CH_2), 31.5 (CH_2), 34.1 (CH_2), 127.8 (CH), 128.0 (CH), 129.9 (CH), 130.1 (CH), 180.6 (C = 0)。该化合物为不饱和脂肪酸。

Hexadecanoic acid (2): 无色针状结晶, $C_{16}H_{32}O_2$ 1H NMR ($CDCl_3$, 400 MHz) δ_H : 2.34 (2H, t, J = 7.5 Hz, H₂-2), 1.62 (2H, brt, J = 7.3 Hz, H₂-3), 1.30~1.25 (24H, brs, 12 × - CH_2), 0.88 (3H, t, J = 6.5 Hz, Me-16); ^{13}C NMR ($CDCl_3$, 100 MHz) δ_C : 180.4 (C-1), 34.1 (C-2), 24.9 (C-3), 29.7~29.0 (10 × - CH_2), 31.9 (C-14), 22.7 (C-15), 14.1 (C-16)。由波谱数据推出该化合物为不饱和脂肪酸。

Senkirkine (3): 无色油状, $C_{19}H_{27}NO_6$ 1H NMR ($CDCl_3$, 400 Hz): δ_H 6.09 (1H, t, J = 2.3 Hz, H-2), 3.32 (1H, br d, J = 18, H-3a), 3.16 (1H, ddd, J = 18, 2.6, 2.3 Hz, H-3b), 2.76 (1H, ddd, J = 12.7, 5.7, 4.2 Hz, H-5a), 2.65 (1H, ddd, J = 12.7, 12.0, 4.2 Hz, H-5b), 2.46 (1H, dddd, J = 11.4, 12.0, 5.7, 3.2 Hz, H-6a), 2.28 (1H, dq, J = 14.4, 4.2 Hz, H-6b), 4.92 (1H, dd, J = 4.4, 3.2 Hz, H-7), 5.32 (1H, d, J = 11.4, Hz, H-9a), 4.38 (1H, br d, J = 11.4, H-9b), 1.76 (1H, ddq, J = 11.0, 2.5, 7.1 Hz, H-13), 2.12 (1H, dd, J = 14.0, 11.0 Hz, H-14a), 1.27 (s, H-18), 0.84 (3H, d, J = 7.1, H-19), 6.54 (1H, t, J = 6.8, H-20), 4.71 (1H, dd, J = 14.5, 6.8 Hz, Ha-2l), 4.60 (1H, dd, J = 14.5, 6.8 Hz, Hb-2l), 2.04 (3H, s, H-22); ^{13}C NMR ($CDCl_3$, 100 MHz) δ_C : 134.5 (s, C-1), 137.0 (d, C-2), 58.9 (t, C-3), 53.5 (t, C-5), 36.2 (t, C-6), 78.0 (d, C-7), 64.0 (t, C-9), 178.0 (s, C-11), 76.6 (s, C-12), 38.5 (t, C-13), 37.6(t, C-14), 134.3 (s, C-15), 166.4 (s, C-16), 24.5 (q, C-18), 10.9 (q, C-19), 137.0 (d, C-20), 58.9 (t, C-21), 40.6 (q, C-22)。由波谱数据推出, 该化合物为吡咯烷生物碱。

(-)-cryptomerion (4): 无色油状, $C_{15}H_{22}O$, ESI-MS m/z 218 [M]+; 1H NMR(600 MHz, $CDCl_3$) δ_H : 6.75 (1H, m, H-3), 2.59 (1H, ddd, J = 1.5, 3.7, 15.9 Hz, Ha-4), 2.35 (1H, dd, J = 13.2, 15.9 Hz, Ha-4), 2.69 (1H, ddd, J = 4.0, 10.2, 13.6 Hz, H-5), 2.46 (1H, dtm, J = 4.4, 18.3 Hz, Ha-6), 2.27 (1H, ddm, J = 11.0, 18.3 Hz, Hb-6), 2.05 (2H, m, H-8), 2.11 (2H, m, H-9), 5.09 (1H, tq, J = 1.5, 6.8 Hz, H-10), 1.61 (3H, s, H3-12), 1.68 (3H, d, J = 1.0 Hz H3-13), 4.85 (1H, s, Ha-14), 4.82 (1H, s, Hb-14), 1.79 (3H, dt, J = 1.4, 2.4 Hz, H3-15); ^{13}C NMR (150 MHz, $CDCl_3$) δ_C : 199.8 (s, C-1), 135.3 (s, C-2), 144.6 (d, C-3), 31.4 (t, C-4), 41.1 (d, C-5), 43.5 (t, C-6), 150.7 (s, C-7), 34.2 (t, C-8), 26.6 (t, C-9), 123.7 (d, C-10), 131.9 (s, C-11), 25.6 (q, C-12), 17.7 (q, C-13), 109.1 (t, C-14), 15.7 (q, C-15)。

2,2-dimethyl-6-acetylchromanone (5): 无色油状, $C_{13}H_{14}O_3$, ESI-MS m/z 218 [M]+; 1H NMR (400 MHz, methanol-d4) δ_H : 1.49 (6H, s, $CH_3 \times 2$), 2.59 (3H, s, - $COCH_3$), 2.77 (2H, s, H-2), 7.00 (1H, d, J = 8.8 Hz, H-8), 8.13 (1H, dd, J = 8.8, 2.2 Hz, H-7), 8.45(1H, d, J = 2.2 Hz, H-5); ^{13}C NMR (100 MHz, methanol-d4) δ_C : 80.3

(s, C-1), 48.5 (t, C-2), 191.7 (s, C-3), 130.2 (s, C-4), 135.4 (d, C-5), 119.1 (d, C-6), 128.1 (d, C-7), 119.0 (d, C-8), 163.4 (s, C-9), 26.6 ($\text{CH}_3 \times 2$), 26.4 (- COCH_3), 196.3 (- COCH_3)。

β -oploplonenone (6): 无色油状; $\text{C}_{15}\text{H}_{24}\text{O}$, ESI-MS m/z 220 [M]+; ^1H NMR (400 MHz, methanol-d4) δ_H : 1.34 (2H, m, H-1), 1.62 (1H, dtd, $J = 20.0, 7.4, 12.8$ Hz, H-2a), 1.81 (1H, m, H-2b), 1.77 (1H, m, H-3), 1.25 (1H, m, H-4), 1.34 (1H, m, H-5), 1.10 (1H, dq, $J = 4.4, 13.0$ Hz, H-6a), 1.75 (1H, m, H-6b), 1.95 (1H, m, H-7a), 2.35 (1H, ddd, $J = 2.4, 4.3, 13.4$ Hz, H-7b), 1.83 (1H, m, H-9), 4.54 (1H, q, $J = 1.7$ Hz, H-10a), 4.62 (1H, q, $J = 1.7$, H-10b), 1.95 (1H, m, H-11), 0.95 (3H, d, $J = 6.9$ Hz, H3-12), 0.78 (3H, d, $J = 6.9$ Hz, H3-13), 1.18 (3H, q, H3-15); ^{13}C NMR (100 MHz, methanol-d4) δ_C : 28.5 (t, C-1), 27.7 (t, C-2), 50.4 (d, C-3), 52.9 (d, C-4), 53.5 (d, C-5), 29.7 (t, C-6), 36.3 (t, C-7), 152.2 (s, C-8), 56.8 (d, C-9), 103.9 (t, C-10), 30.9 (d, C-11), 22.3 (q, C-12), 16.1 (q, C-13), 214.7 (s, C-14), 29.7 (q, C-15)。

7β -(3-ethyl-cis-crotonoyloxy)-14-hydroxy-notonipetranone (7): 无色油状; $\text{C}_{26}\text{H}_{46}\text{O}_6$, ESI-MS m/z 448 [M]+; ^1H NMR (400 MHz, methanol-d4) δ_H : 5.46 (1H, d, $J = 4.0$ Hz, H-1 β), 5.54 (1H, br s, H-7 α), 4.78 (1H, s, Ha-10), 5.18 (1H, s, Hb-10), 2.37 (1H, m, H-11), 1.00 (3H, d, $J = 6.6$ Hz, H3-12), 0.82 (3H, t, $J = 7.0$ Hz, H3-13), 1.23 (3H, d, $J = 6.6$ Hz, H3-15), 5.61 (1H, s, H-2'), 2.16 (1H, d, $J = 7.7$ Hz, H-4'), 1.07 (3H, t, $J = 7.3$ Hz, H3-5'), 2.10 (3H, s, H3-6'), 2.37 (1H, m, H-2''), 0.89 (3H, t, $J = 7.5$ Hz, H3-4''), 1.15 (3H, d, $J = 6.6$ Hz, H3-5''); ^{13}C NMR (100 MHz, methanol-d4) δ_C : 73.2 (d, C-1), 206.2 (s, C-2), 60.4 (d, C-3), 47.3 (d, C-4), 45.2 (d, C-5), 30.2 (t, C-6), 74.2 (d, C-7), 142.6 (s, C-8), 41.7 (d, C-9), 113.0 (t, C-10), 28.8 (d, C-11), 21.7 (q, C-12), 17.0 (q, C-13), 68.5 (d, C-14), 15.7 (q, C-15), 175.3 (s, C-1'), 115.4 (d, C-2'), 162.3 (s, C-3'), 34.1 (t, C-4'), 12.2 (q, C-5'), 17.0 (q, C-6'), 165.8 (s, C-1''), 45.2 (d, C-2''), 27.3 (t, C-3''), 11.8 (q, C-4''), 17.0 (q, C-5'')。

14-acetoxy-7 β -(3-ethyl-cis-crotonoyloxy)-1 α -(2-methylbutyryloxy)-notonipetranone (8): 无色油状; $\text{C}_{28}\text{H}_{42}\text{O}_7$, ESI-MSm/z 490 [M]+; ^1H NMR (400 MHz, methanol-d4) δ_H : 5.46 (1H, d, $J = 4.0$ Hz, H-1 β), 5.54 (1H, br s, H-7 α), 4.78 (1H, s, Ha-10), 5.18 (1H, s, Hb-10), 2.37 (1H, m, H-11), 1.00 (3H, d, $J = 6.6$ Hz, H3-12), 0.82 (3H, t, $J = 7.0$ Hz, H3-13), 1.23 (3H, d, $J = 6.6$ Hz, H3-15), 5.61 (1H, s, H-2'), 2.16 (1H, d, $J = 7.7$ Hz, H-4'), 1.07 (3H, t, $J = 7.3$ Hz, H3-5'), 2.10 (3H, s, H3-6'), 2.37 (1H, m, H-2''), 0.89 (3H, t, $J = 7.5$ Hz, H3-4''), 1.15 (3H, d, $J = 6.6$ Hz, H3-5''); ^{13}C NMR (100 MHz, methanol-d4) δ_C : 74.0 (d, C-1), 209.8 (s, C-2), 57.5 (d, C-3), 47.4 (d, C-4), 45.2 (d, C-5), 30.3 (t, C-6), 75.0 (d, C-7), 142.7 (s, C-8), 42.5 (d, C-9), 113.7 (t, C-10), 28.7 (d, C-11), 21.8 (q, C-12), 17.3 (q, C-13), 70.7 (d, C-14), 15.7 (q, C-15), 172.4 (s, C-1'), 115.5 (d, C-2'), 163.6 (s, C-3'), 34.6 (t, C-4'), 12.0 (q, C-5'), 17.3 (q, C-6'), 176.6 (s, C-1''), 45.5 (d, C-2''), 27.8 (t, C-3''), 12.4 (q, C-4''), 16.3 (q, C-5'')。

tussilagone (9): 无色结晶; $\text{C}_{23}\text{H}_{34}\text{O}_5$, ESI-MS m/z 390 [M]+; ^1H NMR (400 MHz, CDCl_3) δ_H : 2.15 (1H, dd, $J = 16.9, 13.9$ Hz, H-1 α), 2.40 (1H, ddd, $J = 16.9, 1.0, 5.5$ Hz, H-1 β), 2.50 (1H, dd, $J = 3.0, 11.0$ Hz, H-3), 1.47 (1H, m, H-4), 1.97 (1H, dddd, $J = 2.0, 2.0, 11.0, 14.0$ Hz, H-5), 1.45 (1H, ddd, $J = 2.0, 11.0, 14.0$ Hz, H-6 α), 2.08 (1H, dt, $J = 2.0, 14.0$ Hz, H-6 β), 5.58 (1H, t, $J = 3.0$ Hz, H-7), 2.60 (1H, dddd, $J = 2.0, 2.0, 5.9$ 11.5, 13.9 Hz, H-9), 5.15 (1H, s-like, H-10Z), 4.79 (1H, d, $J = 1.0$ Hz, H-10E), 2.30 (1H, dqq, $J = 3.0, 6.8, 6.9$ Hz, H-11), 0.99 (3H, d, $J = 6.5$ Hz, H3-12), 0.78 (3H, d, $J = 7.0$ Hz, H3-13), 5.10 (1H, dq, $J = 3.2, 6.6$ Hz, H-14), 1.22 (3H, d, $J = 6.5$ Hz, H3-15), 5.63 (1H, qt, $J = 1.3, 1.3$ Hz, H-2'), 2.17 (2H, dq, $J = 1.3, 7.5$ Hz, H-4'), 1.07 (3H, t, $J = 7.5$ Hz, H3-5'), 2.15 (3H, d, $J = 1.3$ Hz, H3-6'), 2.15 (3H, s, OCOCH_3); ^{13}C NMR (100 MHz, CDCl_3) δ_C : 42.5 (t, C-1), 214.9 (s, C-2), 57.2 (d, C-3), 49.0 (d, C-4), 43.8 (d, C-5), 31.1 (t, C-6), 72.9 (d, C-7), 146.0 (s, C-8), 42.2 (d, C-9), 110.1 (t, C-10), 27.5 (d, C-11), 21.5 (q, C-12), 15.4 (q, C-13), 69.5 (d, C-14),

15.3 (q, C-15), 165.9 (s, C-1'), 114.5 (d, C-2'), 162.0 (s, C-3'), 33.8 (t, C-4'), 11.9 (q, C-5'), 18.9 (q, C-6'), 170.9 (OCOCH₃), 21.3 (OCOCH₃)。

7 β -(3-ethyl-cis-crotonoyloxy)-1 α -(2-methylbutyryloxy)-3(14)-dehydro-Z-notonipetran (10): 无色油状; C₂₆H₃₈O₅, ESI-MS m/z 430 [M]+; 1H NMR (500 MHz, methanol-d4) δ_H : 5.56 (1H, d, J = 4.0 Hz, H-1 β), (1H, d, J = 3.3 Hz, H-7 α), 4.82 (1H, s, Ha-10), 5.18 (1H, s, Hb-10), 2.02 (1H, m, H-11), 0.98 (3H, d, J = 6.6 Hz, H3-12), 0.90 (3H, t, J = 7.0 Hz, H3-13), 6.42 (1H, q, J = 7.3 Hz, H-14), 2.18 (3H, d, J = 7.3 Hz, H3-15), 5.64 (1H, s, H-2'), 1.07 (3H, t, J = 7.3 Hz, H3-5'), 2.15 (3H, s, H3-6'), 2.40 (1H, m, H-2''), 0.87 (3H, t, J = 7.3 Hz, H3-4''), 1.13 (3H, d, J = 7.0 Hz, H3-5''); ¹³C NMR (125 MHz, methanol-d4) δ_C : 73.2 (d, C-1), 200.1 (s, C-2), 139.0 (s, C-3), 44.6 (d, C-4), 40.3 (d, C-5), 29.6 (t, C-6), 72.3 (d, C-7), 140.4 (s, C-8), 45.7 (d, C-9), 112.5 (t, C-10), 27.3 (d, C-11), 21.2 (q, C-12), 15.0 (q, C-13), 136.7 (d, C-14), 15.5 (q, C-15), 165.8 (s, C-1'), 114.4 (d, C-2'), 162.0 (s, C-3'), 33.7 (t, C-4'), 11.4 (q, C-5'), 18.8 (q, C-6'), 175.4 (s, C-1''), 40.8 (d, C-2''), 26.7 (t, C-3''), 11.8 (q, C-4''), 16.5 (q, C-5'')。

(1R,3R,4R,5S,6S)-1,5-diacetoxy-8-angeloyloxy-3,4-epoxybisabola-7(14),10-dien-2-one (11): 无色油状; C₂₄H₃₂O₈, ESI-MS m/z 448 [M]+; 1H NMR (400 MHz, CDCl₃) δ_H : 5.68 (1H, d, J = 12.7 Hz, H-1), 3.40 (1H, s, H-4), 5.35 (1H, d, J = 8.8 Hz, H-5), 2.87 (1H, d, J = 8.8, 12.7 Hz, H-6), 5.23 (1H, dd, J = 3.2, 7.1 Hz, H-8), 2.33 (2H, m, H-9), 5.04 (1H, tq, J = 1.0, 7.3 Hz, H-10), 1.68 (3H, d, J = 1.0 Hz, H3-12), 1.62 (3H, s, H3-13), 5.24 (1H, s, Ha-14), 5.33 (1H, d, J = 0.7 Hz, Hb-14), 1.48 (3H, s, H3-15), 6.08 (1H, qq, J = 1.5, 7.1 Hz, H-3'), 1.97 (3H, dq, J = 1.5, 7.1 Hz, H-4'), 1.88 (3H, dq, J = 1.5, 1.5 Hz, H3-5'); ¹³C NMR (100 MHz, CDCl₃) δ_C : 72.3 (d, C-1), 199.1 (s, C-2), 61.0 (d, C-3), 65.4 (d, C-4), 74.5 (d, C-5), 48.1 (d, C-6), 145.7 (s, C-7), 75.6 (d, C-8), 31.5 (t, C-9), 119.3 (d, C-10), 134.0 (s, C-11), 24.4 (q, C-12), 18.7 (q, C-13), 113.3 (t, C-14), 14.6 (q, C-15), 166.4 (s, C-1'), 127.5 (s, C-2'), 138.1 (d, C-3'), 16.6 (q, C-4'), 19.5 (q, C-5'), 19.3 (1-COCH₃), 169.5 (1-COCH₃), 19.2 (5-COCH₃), 170.1 (5-COCH₃)。

通过波谱数据推出, 化合物 6、7、8、9、10 和 11 为 oplopane 型倍半萜, 属于款冬花中一类较为常见的倍半萜。

(1R,3R,4R,5S,6S)-1-acetoxy-8-angeloyloxy-3,4-epoxy-5-hydroxybisabola-7(14),10-dien-2-one (12): 无色油状; C₂₂H₃₀O₇, ESI-MS m/z 406 [M]+; 1H NMR (400 MHz, CDCl₃) δ_H : 5.68 (1H, d, J = 13.7 Hz, H-1), 3.51 (1H, s, H-4), 4.25 (1H, d, J = 8.5 Hz, H-5), 2.56 (1H, d, J = 8.5, 13.7 Hz, H-6), 4.72 (1H, dd, J = 2.7, 8.8 Hz, H-8), 2.18 (1H, m, Ha-9), 2.52 (1H, m, Hb-9), 5.13 (1H, tq, J = 1.0, 7.1 Hz, H-10), 1.70 (3H, d, J = 1.0 Hz, H3-12), 1.64 (3H, s, H3-13), 5.08 (1H, s, Ha-14), 5.20 (1H, d, J = 0.7 Hz, Hb-14), 1.48 (3H, s, H3-15), 6.20 (1H, qq, J = 1.5, 7.3 Hz, H-3'), 2.00 (3H, dq, J = 1.5, 7.3 Hz, H-4'), 1.92 (3H, dq, J = 1.5, 1.5 Hz, H3-5'); ¹³C NMR (100 MHz, CDCl₃) δ_C : 72.2 (d, C-1), 201.9 (s, C-2), 62.5 (d, C-3), 70.2 (d, C-4), 73.8 (d, C-5), 54.5 (d, C-6), 148.3 (s, C-7), 76.8 (d, C-8), 33.3 (t, C-9), 120.6 (d, C-10), 135.4 (s, C-11), 26.0 (q, C-12), 18.1 (q, C-13), 114.3 (t, C-14), 14.7 (q, C-15), 168.4 (s, C-1'), 129.1 (s, C-2'), 140.0 (d, C-3'), 16.0 (q, C-4'), 20.3 (q, C-5'), 20.7 (1-COCH₃), 171.4 (1-COCH₃)。

1 β ,8-bisangeloyloxy-3 α ,4 α -epoxybisabola-7(14),10-dien-2-one (13): 无色油状; C₂₅H₃₄O₆, ESI-MS m/z 430 [M]+; 1H NMR (600 MHz, methanol-d4) δ_H : 5.74 (1H, d, J = 12.8 Hz, H-1), 3.40 (1H, br d, J = 4.4 Hz, H-4), 2.21 (2H, brddd, J = 4.4, 8.0, 15.0 Hz, H-5), 2.78 (1H, ddd, J = 8.0, 11.6, 12.8 Hz, H-6), 5.07 (1H, dd, J = 6.0, 8.0 Hz, H-8), 2.35 (2H, m, H-9), 5.03 (1H, t, J = 6.3 Hz, H-10), 1.64 (3H, brs, H3-12), 1.58 (3H, brs, H3-13), 5.24 (1H, s, Ha-14), 5.09 (1H, s, Hb-14), 1.42 (3H, s, H3-15), 6.05 (1H, qq, J = 1.2, 7.2 Hz, H-3'), 1.95 (3H, dd, J = 1.2, 7.2 Hz, H-4'), 1.86 (3H, d, J = 1.2 Hz, H3-5'), 5.60 (1H, d, J = 1.2 Hz, H-2''), 2.12 (2H, br

q, $J = 7.4$ Hz, H-4''), 1.01 (3H, t, $J = 7.4$ Hz, H3-5''), 2.09 (3H, br s, H-6''); ^{13}C NMR (150 MHz, methanol-d4) δ_{C} : 75.6 (d, C-1), 203.4 (s, C-2), 62.8 (d, C-3), 65.8 (d, C-4), 33.4 (t, C-5), 45.5 (d, C-6), 149.4 (s, C-7), 76.6 (d, C-8), 32.9 (t, C-9), 120.6 (d, C-10), 135.6 (s, C-11), 26.1 (q, C-12), 18.3 (q, C-13), 114.7 (t, C-14), 15.2 (q, C-15), 168.3 (s, C-1'), 129.3 (s, C-2'), 139.7 (d, C-3'), 16.2 (q, C-4'), 21.3 (q, C-5'), 167.0 (s, C-2''), 114.3 (d, C-2''), 164.7 (s, C-3''), 32.9 (t, C-4''), 12.4 (q, C-5''), 19.1 (q, C-6'')。

1 β -(3-ethyl-cis-crotonoyloxy)-8-angeloyloxy-3 α ,4 α -epoxy-bisabola-7(14),10-dien-2-one (14): 无色油状; C25H34O6, ESI-MS m/z 430 [M]+; 1H NMR (400 MHz, methanol-d4) δ_{H} : 5.74 (1H, d, $J = 12.8$ Hz, H-1), 3.40 (1H, br d, $J = 4.4$ Hz, H-4), 2.21 (2H, brddd, $J = 4.4, 8.0, 15.0$ Hz, H-5), 2.78 (1H, ddd, $J = 8.0, 11.6, 12.8$ Hz, H-6), 5.07 (1H, dd, $J = 6.0, 8.0$ Hz, H-8), 2.35 (2H, m, H-9), 5.03 (1H, t, $J = 6.3$ Hz, H-10), 1.64 (3H, brs, H3-12), 1.58 (3H, brs, H3-13), 5.24 (1H, s, Ha-14), 5.09 (1H, s, Hb-14), 1.42 (3H, s, H3-15), 6.05 (1H, qq, $J = 1.2, 7.2$ Hz, H-3'), 1.95 (3H, dd, $J = 1.2, 7.2$ Hz, H-4'), 1.86 (3H, d, $J = 1.2$ Hz, H3-5'), 5.60 (1H, d, $J = 1.2$ Hz, H-2''), 2.12 (2H, br q, $J = 7.4$ Hz, H-4''), 1.01 (3H, t, $J = 7.4$ Hz, H3-5''); ^{13}C NMR (100 MHz, methanol-d4) δ_{C} : 75.4 (d, C-1), 202.9 (s, C-2), 62.7 (d, C-3), 65.7 (d, C-4), 33.6 (t, C-5), 46.5 (d, C-6), 149.3 (s, C-7), 75.6 (d, C-8), 32.1 (t, C-9), 120.5 (d, C-10), 135.5 (s, C-11), 26.0 (q, C-12), 18.1 (q, C-13), 113.5 (t, C-14), 15.1 (q, C-15), 168.1 (s, C-1'), 128.6 (s, C-2'), 139.3 (d, C-3'), 16.1 (q, C-4'), 20.7 (q, C-5'), 168.2 (s, C-2''), 129.0 (s, C-2''), 139.5 (d, C-3''), 16.1 (q, C-4''), 20.8 (q, C-5'')。

(4R,6E)-2-acetoxy-8-angeloyloxy-4-hydroxybisabola-2,6,10-trien-1-one (15): 无色油状; C22H30O6, ESI-MS m/z 390 [M]+; 1H NMR (600 MHz, methanol-d4) δ_{H} : 3.99 (1H, ddd, $J = 8.1, 3.7, 3.7$ Hz, H-5), 2.52 (1H, brddd, $J = 14.7, 3.7$ Hz, H-6 α), 2.96 (1H, dd, $J = 14.7, 3.7$ Hz, H-6 β), 5.54 (1H, t, $J = 7.3$ Hz, H-8), 2.12 (2H, m, H-9), 4.96 (1H, br t, $J = 6.3$ Hz, H-10), 1.54 (3H, s, H3-12), 1.44 (3H, s, H3-13), 1.81 (3H, s, H3-14), 2.11 (3H, d, $J = 1.8$ Hz, H3-15), 5.63 (1H, qq, $J = 7.0, 1.5$ Hz, H-3'), 1.82 (1H, dq, $J = 7.0, 1.5$ Hz, H-4'), 1.71 (3H, dq, $J = 1.5, 1.5$ Hz, H3-5'), 1.89 (3H, s, OCOCH₃); ^{13}C NMR (150 MHz, methanol-d4) δ_{C} : 184.8 (s, C-1), 146.9 (s, C-2), 149.6 (s, C-3), 69.4 (d, C-4), 37.9 (t, C-5), 128.9 (s, C-6), 144.6 (s, C-7), 75.0 (d, C-8), 32.7 (t, C-9), 119.7 (d, C-10), 137.0 (s, C-11), 26.2 (q, C-12), 18.1 (q, C-13), 15.6 (q, C-14), 14.1 (q, C-15), 168.6 (s, C-1'), 128.9 (s, C-2'), 140.1 (d, C-3'), 16.2 (q, C-4'), 20.3 (q, C-5'), 20.9 (1-COCH₃) 170.4 (1-COCH₃)。

通过波谱数据得出, 化合物 12、13、14 和 15 均为没药烷型倍半萜。

5. 结果与讨论

在对款冬花中化学成分的研究中, 发现其含有吡咯烷生物碱[16]、倍半萜[17]、酚类化合物[18]、黄酮[19]等化学成分, 不管是国外学者, 还是国内学者对其成分研究最多的是生物碱、酚类化合物和黄酮。对于款冬花中所含的倍半萜化合物还要追溯于 20 世纪 90 年代末由 Yaoita Y 等分离得到新的没药烷型倍半萜[9] [14], 近年来, 也有学者从款冬花植物中发现了两个新颖结构的 oplopnone 型倍半萜 7 β -angeloyloxy-14-hydroxy-notonipetranone 和 1 α -hydroxy-7 β -(4-methylsenecioyloxy)-oplopone-3(14)Z,8(10)-dien-2-one, 一个新颖结构的没药烷型倍半萜 1 α -(3"-ethyl-cis-crotonoyloxy)-8-angeloyloxy-3 β ,4 β -epoxy-bisabola-7(14),10-diene [3]。本文中共分离得到 20 个化合物, 运用各种波谱手段鉴定得到 15 个化合物, 其中化合物 1 和 2 为脂肪酸, 化合物 3 为吡咯烷生物碱, 化合物 6、7、8、9、10 和 11 为 oplopnone 型倍半萜, 化合物 12、13、14 和 15 均为没药烷型倍半萜。

款冬花中报道的倍半萜具有广泛的活性, 包括抗炎、抗过敏、抗癌、神经保护等。本文通过鉴定化学结构, 探讨结构特征, 从结构特征上看, 款冬花中主要以 oplopnone 型倍半萜和没药烷型倍半萜为主, 其他骨架类型的倍半萜相对较少。而且, 这些结构的化学活性相对较低。此外, 有文献报道的款冬花中

抗炎活性的结果与构效关系有一定关系。因此，我们下一步将对已分离得到的化合物进行抗炎活性的筛选，从而发现其抗炎活性与结构之间的关系。

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参考文献

- [1] 李静, 秦雪梅, 李震宇. 款冬花中倍半萜类成分的研究进展[J]. 中草药, 2017, 48(14): 2964-2971.
- [2] 钟云青. 款冬花散治疗慢性阻塞性肺疾病急性加重期(痰热郁肺证)临床观察[J]. 中国中医急症, 2017, 26(1): 149-151.
- [3] Li, W., Huang, X. and Yang, X.W. (2012) New Sesquiterpenoids from the Dried Flower Buds of *Tussilago farfara* and Their Inhibition on NO Production in LPS-Induced RAW264.7 Cells. *Fitoterapia*, **83**, 318-322. <https://doi.org/10.1016/j.fitote.2011.11.011>
- [4] Jang, H., Lee, J.W., Lee, C., Jin, Q., Choi, J.Y., Lee, D., Han, S.B., Kim, Y., Hong, J.T., Lee, M.K. and Hwang, B.Y. (2016) Sesquiterpenoids from *Tussilago farfara* Inhibit LPS-Induced Nitric Oxide Production in Macrophage RAW 264.7 Cells. *Archives of Pharmacal Research*, **39**, 127-132. <https://doi.org/10.1007/s12272-015-0667-7>
- [5] Liu, L.L., Yang, J.L. and Shi, Y.P. (2011) Sesquiterpenoids and Other Constituents from the Flower Buds of *Tussilago farfara*. *Journal of Asian Natural Products Research*, **13**, 920-929. <https://doi.org/10.1080/10286020.2011.600251>
- [6] 刘世旺, 付宏征, 林文翰. 糙苏的化学成分研究(I) [J]. 中草药, 1999(3): 161-164.
- [7] Shen, Q., Cai, G.M., He, G.X., et al. (2009) Study on the Chemical Constituents of Fruits of *Cannabis sativa* L. *Natural Product Research and Development*, **21**, 784-786.
- [8] Roxana, L., Andrea, S., Susanne, S., Christine, R., Liselotte, K. and Brigitte, K. (2000) Quantitative Analysis of the Pyrrolizidine Alkaloids Senkirkine and Senecionine in *Tussilago farfara* L. by Capillary Electrophoresis. *Phytochemical Analysis*, **11**, 366-369. [https://doi.org/10.1002/1099-1565\(200011/12\)11:6<366::AID-PCA538>3.0.CO;2-1](https://doi.org/10.1002/1099-1565(200011/12)11:6<366::AID-PCA538>3.0.CO;2-1)
- [9] Yaoit, Y., Suzuki, N. and Kikuchi, M. (2001) Structures of New Sesquiterpenoids from Farfarae Flos. *Chemical and Pharmaceutical Bulletin*, **49**, 645-648. <https://doi.org/10.1248/cpb.49.645>
- [10] Liu, Y.F., Yang, X.W. and Wu, B. (2007) Chemical Constituents of the Flower Buds of *Tussilago farfara*. *Journal of Chinese Pharmaceutical Sciences*, **16**, 288-293.
- [11] Piers, E. and Gavai, A.V. (1990) A (Z)-Ethylidenecyclopentane Annulation Method. Total Syntheses of (\pm)-Anhydrooplopanone, (\pm)-Oplopanone, and (\pm)-8-*epi*-Oplopanone. *The Journal of Organic Chemistry*, **55**, 2380-2390. <https://doi.org/10.1021/jo00295a028>
- [12] Kikuchi, M. and Suzuki, N. (1992) Studies on the Constituents of *Tussilago farfara* L. II. Structures of New Sesquiterpenoids Isolated from the Flower Buds. *Chemical and Pharmaceutical Bulletin*, **40**, 2753-2755. <https://doi.org/10.1248/cpb.40.2753>
- [13] Park, H.R., Yoo, M.Y., Seo, J.H., Kim, I.S., Kim, N.Y., Kang, J.Y., Cui, L., Lee, C.S., Lee, C.H. and Lee, H.S. (2008) Sesquiterpenoids Isolated from the Flower Buds of *Tussilago farfara* L. Inhibit Diacylglycerol Acyltransferase. *Journal of Agricultural and Food Chemistry*, **56**, 10493-10497. <https://doi.org/10.1021/jf801978r>
- [14] Yaoit, Y., Suzuki, N. and Kikuchi, M. (2001) Structures of New Sesquiterpenoids from Farfarae Flos. *Chemical and Pharmaceutical Bulletin*, **49**, 645-648. <https://doi.org/10.1248/cpb.49.645>
- [15] Baba, H., Yaoita, Y. and Kikuchi, M. (2007) Sesquiterpenoids from *Ligularia dentata*. *Helvetica Chimica Acta*, **90**, 1302-1312. <https://doi.org/10.1002/hlca.200790131>
- [16] Editorial Committee of Flora of China. Chinese Academy of Sciences (1999) Flora of China. Vol. 77(1), Science Press, Beijing, 93.
- [17] Xue, S.Y., Li, Z.Y., Zhi, H.J., Sun, H.F., Zhang, L.Z., Guo, X.Q. and Qin, X.M. (2012) Metabolic Fingerprinting Investigation of *Tussilago farfara* L. by GC-MS and Multivariate Data Analysis. *Biochemical Systematics and Ecology*, **41**, 6-12. <https://doi.org/10.1016/j.bse.2011.11.003>
- [18] Akçin, Ö.E. (2007) Morphological and Anatomical Characteristics of *Cichorium intybus* L., *Tragopogon latifolius* Boiss. and *Tussilago farfara* L. (Asteraceae). *International Journal of Natural and Engineering Sciences*, **1**, 81-85.
- [19] 王建全, 张亚娜, 林敏. 乙醇/硫酸铵双水相体系分离纯化款冬花总黄酮[J]. 井冈山大学学报(自然科学版), 2016, 37(1): 40-45.